STUDY THE MICROPLASTIC ACCUMULATE BACTERIAL PATHOGEN SYNTHESIS AND CHARACTERISATION IN ECOSYSTEM

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Abstract

The buildup of plastic waste is becoming a more and more pressing issue for marine areas. The goal of this study is to find a cost-effective and efficient method for determining the concentrations of microplastics in marine sediments. If the opportunity arises during the project's routine benthic fauna surveys, this will be incorporated into the process. Microplastic debris greater than one millimetre in diameter was counted and categorised as part of a routine monitoring of the benthic ecology in twenty-two separate Marine Protected Areas (MPAs) in English inshore seas. Using samples from the places where the samples were gathered, this was done. The mean density of microplastic particles per 0.1 m2 ranged from 0.2 in the Dover to Deal Marine Conservation Zone to 42.7 in the Mersey Estuary Special Protection Area. 61.2 percent of the samples were found to have microplastic particles. One or two locations that were located closer to cities or industrialised areas had considerable levels of plastic, as did several of the more distant sites. To tackle marine plastic pollution, which should be dealt with upstream, at its source, spatial protection measures such as marine protected areas (MPAs) are not an acceptable tool in and of themselves. These regions are referred to as "marine protected areas" because of this (MPAs).

Introduction

Plastic waste, an inevitable and inadvertent marker of the Anthropocene, has become a ubiquitous pollutant in nature. Plastics can therefore exert negative efects on biota in both, aquatic and terrestrial ecosystems. Direct consequences of larger plastic waste for organisms range from entanglement and sufocation to intestinal obstruction. Microplastic (MP) particles (the most common form of fragments, are taken up by various vertebrate and invertebrate species, leading to extensive bioaccumulation. By adsorbing a multitude of hydrophobic organic substances, plastic solid waste (PSW) may form an eco-corona and thus, inter alia, interfere with chemical communication in aquatic systems and biomagnify potentially hazardous xenobiotics in the food web. Humans are constantly exposed to MP

through ingestion, inhalation, and skin contact. Internalised particles can, for instance, cause respiratory infammation, lung disease and endocrinological disorders due to a combination of intrinsic toxicity and chemical leaching. A trending, yet poorly understood aspect of plastic pollution with potential efects on ecosystems and human health is the interaction between plastics and microbes. Te hydrophobic surface of plastic waste provides an ideal environment for microbial colonisation and bioflm formation, and represents a protective ecological niche, the so-called 'plastisphere. Tese epiplastic communities harbour Archaea and Bacteria as well as unicellular and oligocellular eukaryotes including fungil, and have been found on plastics from marine, limnic, and fuvial ecosystems in numerous biomes from the equator to the polar regions. Metagenomic studies show that MP selects for microbial communities that are different from the surrounding environment19, and whose composition and succession is subject to spatial and seasonal infuence as well as polymer type. Tus, MP represents a microhabitat with a high selectivity and plasticity, which can have efects at ecosystem level, such as the spread of antibiotic resistances through the concentration of certain lineages or changes in microbial nitrogen and carbon cycle dynamics through shifs in community structure . In this context, the role of MP as a reservoir and vector for invasive and harmful microbes is a recurring aspect in the relevant literature. Te durable substrates can not only massively promote adhesion, thus serving as reservoirs for pathogens, but can also be transported over long distances by wind, currents, and waves, eventually leading to the establishment of alien communities at specifc destinations. In marine surface waters, foating PSW acts as vector for the distribution of potentially harmful bacteria of the genus Vibrio and pathogenic serotypes of Escherichia coli25 as well as invasive algal species. In addition, marine PSW has been reported to harbour microbial pathogens that can cause disease outbreaks in coral reefs, fsh, and shellfsh. Although research on the plastisphere and its ecological impact has made enormous progress in recent years, we are far from an integral understanding. Reasons for the limited state of knowledge on the plastisphere holobiome are, on the one hand, the focus on prokaryotic communities, and, on the other hand, the somewhat limited consideration of terrestrial ecosystems. Fungi are the ideal group of organisms for studying microbial plastic colonisation in terrestrial systems, as they are particularly well adapted to life in the plastisphere due to their adsorptive nutrition mode, apical growth, invasive growth forms, bioflm formation, and the secretion of hydrophobic proteins (hydrophobins). Most phylogenetically higher (non-zoosporic) fungi are not bound to the aqueous phase for their propagation, produce far more biomass in certain soils than prokaryotes, and in principle can therefore systematically colonise soil-deposited plastic waste. Fungi are early colonisers of drifing MP, part of polymicrobial bioflms e.g., on domestic plastic surfaces, and have also been isolated from landfll plastics. Pathogenic fungal taxa such as Candida, Fusarium and Rhodotorula are known to occur on plastic surfaces of medical, industrial and household appliances. Tus, MP potentially can play a role in the accumulation and spread of fungal pathogens in soil environments receiving massive infux of PSW, such as home gardens, roadsides, agricultural soils, and landflls. So far, studies on such efects of MP have been carried out in aquatic systems and remote areas without considering the immediate human environment. We addressed these knowledge gaps by providing frst indepth insights into fungal communities of the plastisphere biome and by evaluating the role of MP as a carrier of potentially pathogenic fungi in terrestrial ecosystems. Our three hypotheses were that soil-deposited MP (1) is readily colonised by fungal bioflms, (2) hosts a distinct mycobiome different from that of the surrounding bulk soil, and (3) accumulates a

variety of pathogenic species, including opportunistic human pathogens. We conceptualised an operational design allowing the comparative study of soil-inhabiting and plastic-associated assemblages in situ in human settings. Terefore, we collected fve biological replicates from the topsoil of fve diferent sites with high human activity and highlevel plastic pollution within the municipal boundary of Siaya, Western Kenya. Te sites included two landflls, a marketplace, a roadside, and a courtyard. ITS metabarcoding was applied to decipher the fungal community diversity of the plastic and soil (sub)samples obtained through selective subsampling, while scanning electron microscopy (SEM) and confocal laser scanning microscopy (CLSM) were used to visualise patterns of fungal plastic colonisation. Finally, we used trait data from multiple sources (meta-analysis) to construct a functional profle of the dominant fungal phylotypes of the plastisphere. From these findings, we demonstrate the role of MP as a selective microhabitat and 'hot spot' for potentially human pathogenic fungal species in terrestrial ecosystems in general, and the immediate human environment in sub-Saharan Africa in particular.

Related work

(Sudarsono, Sudarsono, et al., 2017) The pathogen causing Phalaenopsis soft rot disease and developed detached leaf inoculation methods were identified. Based on its 16S rDNA sequences, the pathogen causing soft rot disease in Phalaenopsis was Erwinia chrysanthemi/Dickeya chrysanthemi. Both virulent and avirulent strains were revealed. The detached leaf inoculation assay for E. chrysanthemi/D. chrysanthemi resistance evaluation included wounding and inoculating the detached leaf with 108 CFU/ml of bacteria. Soft rot disease symptoms in the inoculated detached leaf were measurable at 20 h after inoculation. The detached leaf assay was applicable for evaluating Phalaenopsis germplasm and progeny resistance in Phalaenopsis breeding programs.

(Pagter, Elena, et al. , 2020) Plastic pollution is prevalent in all habitats and microplastic ingestion has been recorded in several different species examined to date. However, most studies have focused solely on commercial species. This study investigates microplastics (MPs) by assessing the levels present in a mixed demersal trawl at two sites in a coastal embayment. MPs were recovered from species' gastrointestinal tracts and polymers identified with μ FTIR spectroscopic analysis. Particles recovered comprised 20% natural fibres. The majority of MPs were identified as PE, PVDF, and PETE. Results show an average MP range of 0.11-4.67 MPs individual⁻¹. Fluctuating trendlines for MPs within species suggest that their bioavailability is influenced by several factors. Individual species show significant differences in ingested MP between trawls; however, when the entire trawl community is assessed there is no significant difference between sites.

(Yusof Shuaib Ibrahim et al., 2017) The presence of microplastics (<1 mm) in wild and cagecultured Asian sea bass (Lates calcarifer) was successfully studied. Fish samples were collected from Setiu Wetlands in October 2016. Microplastics were isolated from fish samples using the alkaline solution method (10 M of NaOH solution). Microplastics were sorted visually according to their shapes and colours after being observed under dissecting microscope. A total of 4,498 pieces of microplastics were identified and threadlike shape was the most abundant microplastic particles found during this study. (Maughan, H., et al. 2010)ptake signal sequences are DNA motifs that promote DNA uptake by competent bacteria in the family Pasteurellaceae and the genus Neisseria. The genomes of these bacteria contain many copies of their canonical uptake sequence (often >100-fold overrepresentation), so the bias of the uptake machinery causes cells to prefer DNA derived from close relatives over DNA from other sources. However, the molecular and evolutionary forces responsible for the abundance of uptake sequences in these genomes are not well understood, and their presence is not easily explained by any of the current models of the evolution of competence. Here we describe use of a computer simulation model to thoroughly evaluate the simplest explanation for uptake sequences, that they accumulate in genomes by a form of molecular drive generated by biased DNA uptake and evolutionarily neutral (i.e., unselected) recombination. In parallel we used an unbiased search algorithm to characterize genomic uptake sequences and DNA uptake assays to refine the Haemophilus influenzae uptake specificity.

(Brear, Paul, et al., 2012) An essential feature of nanotechnology is the search for more environmentally friendly methods of producing copper nanoparticles (CuNPs). Recent years have seen the development of a novel approach to the synthesis of various nanoparticles. This approach involves the utilisation of secondary metabolites derived from aqueous plant leaf extract. After that, copper nanoparticles were produced by using NEEM (Neem extract water), and further study aimed at improving the conditions in which these nanoparticles might be produced. The phytochemicals found in the plant are responsible for the reduction of Cu2+ ions to CuNPs, in addition to performing the functions of a capping and stabilising agent. During the manufacturing process of CuNPs, researchers were able to monitor their progress by analysing the absorbance spectrum. Studies conducted with ATR-FTIR, SEM, HRTEM, and XRD all found evidence of the presence of CuNPs.

(Green, Benjamin C. et al., 2019) The accumulation of trash made of plastic is becoming an increasingly urgent issue for marine ecosystems. As an opportunistic addition to the routine sampling of benthic infauna, the purpose of this project is to investigate a method that is both cost-effective and efficient for quantifying larger proportions of microplastics found in marine sediments. In the course of the routine monitoring of the benthic ecosystem in twenty-two different Marine Protected Areas in English inshore seas, a subsample of microplastics larger than one millimetre in size was counted and categorised from the sediment samples that were collected. There was evidence of microplastic particles in 61.2% of the samples that were taken, with the mean density of these particles per 0.1 m2 varying from 0.2 in the Dover to Deal MCZ to 42.7 in the Mersey Estuary Special Protection Area. At some of the more distant sites, as well as some that were closer to urban or industrialised regions, researchers found high quantities of plastic. In and of themselves, spatial protection measures like marine protected areas (MPAs) are not an appropriate instrument for combating marine plastic pollution, which rather to be tackled upstream at its source.

Proposed methodology

The extraction of DNA from a wide variety of bacterial species. Table 1, which can be seen on this page, provides a comprehensive summary of the findings obtained from the investigation into the various bacterial strains. Every strain was cultivated in either a broth or on agar plates that were supplemented with glucose as a nutrient. The glucose served as a source of nutrients in both of these different mediums. In order to successfully prepare the template for the PCR amplification, whole bacterial cells were used as the source material. In order to achieve lysis, one loopful of cells from a colony that had been resuspended in one hundred litres of sterile distilled water were subjected to five minutes of heating at a temperature of one hundred degrees Celsius inside of a thermal cycler model PTC-200. This was done in order to eliminate any microorganisms that might have been present (MJ Research). After chilling the culture for five minutes at 0 degrees Celsius, five microliters of the prepared culture were used in each response. The culture was first chilled for five minutes. There was no other treatment that was used.

a method that is typically known as selective subsampling is an example of a strategy. To begin, using tweezers that had been thoroughly cleaned, any non-soil particles that were visible in the initial sample were removed and placed in a separate container. This was done so that the soil could be examined more closely. After removing 100 mg of soil material from each sample with a sterile spatula and placing it in a screw-cap tube, the tube was then weighed on a fne balance to determine whether or not it should be considered a soil subsample. The results of this weighing helped determine whether or not the soil subsample should be considered a soil subsample. This was successfully completed. While this was going on, the fragments that had been separated were visually characterised by using a Stemi SV 11 stereo microscope (Zeiss, Oberkochen, Germany). This was done so that various types of plastic particles could be differentiated from one another. In order to generate a comparable plastic subsample, first 100 mg of small plastic particles were deposited in a tube, and then the tube was given a thorough washing twice with sterile deionized water in order to remove any connected soil particles. Finally, the tube was analysed in order to generate the comparable plastic subsample. Following this, the little pieces of plastic were given a second thorough washing with sterile deionized water (larger fragments were intentionally disregarded). One hundred pieces of plastic were selected at random from each location and sample, and then those pieces were measured so that an accurate representation of the size distribution could be produced. A stereo microscope was utilised in order to acquire these values for measurement. Around 78 percent of the particles had a size that was between 3 millimetres and 30 millimetres, and the diameters of the particles ranged from approximately 3 millimetres to 30 millimetres.

DNA that was extracted from bacteria that used plants as their primary source of supply material. Testing with PCR was carried out in order to identify whether or not Cmi was present in the stems and roots of inoculated lucerne plants. After being sliced into thin cross sections with sharp blades, the samples were mashed with a mortar and pestle before being placed in a thermal cycler model PTC-200 and heated for five minutes at a temperature of one hundred degrees Celsius. This process was repeated three times (MJ Research). After chilling the culture for five minutes at 0 degrees Celsius, five microliters of the prepared culture were used in each response. The culture was first chilled for five minutes. There was no other treatment that was used.

Polymerase chain reaction is referred to as P.C.R. (polymerase chain reaction) for short. Primer concentrations were 0.5, 2, 0.2, and 1.2 mM for the primers, while the deoxynucleoside triphosphates were 0.2 mM each. Other components were 1x PCR buffer made by Promega, 1.5 mM MgCl2, and 2 units of Promega Taq DNA polymerase. To conduct the PCR, we used a MJ Research PTC-200 thermal cycler. We ran one cycle at 94°C for 3 minutes, followed by 61°C for 30 seconds, and 72°C for 2 minutes of elongation. We

then ran 40 cycles with the following parameters: 94°C for 30 seconds, 61°C for 30 seconds, and 72°C for 2 minutes of elongation.

We're doing a check on you. Based on histopathological symptoms in root cross-sections, the disease severity of infected lucerne plants was graded on a 7-point scale. In the xylem, little reddish brown or dark brown spots can be found in the centre; 2–yellowing in the centre; 3–a half-yellowish ring around the perimeter; 4–a continuous yellow band; 5–darkening of the yellow band; and 6–a total yellowing of the xylem or death of the entire plant. It is scored on a scale of 0–6, with zero discolouration being the worst.

Various plant pathogenic and saprophytic bacteria were found in samples 3–30, including DNA from Cmm, Cmn, and Cf (Table 1, Figure 1). A PCR test confirmed the presence of Cmi in the roots and stems of plants injected with the bacteria that causes the sickness (Table 2). To determine whether or not the inoculations were successful, PCR was used to test plants that had already been injected but showed no signs of disease.

Clavibacter michiganensis, subsp. insidiosus was found in Lucerne plants infected with infected soil

| | | PCR | | Microbiological analysis | |
|-----------------------------------|---|-------|-------|--------------------------|-------|
| | | stems | roots | stems | roots |
| Plants with histological symptoms | 0 | - | - | - | - |
| | 1 | - | + | - | + |
| | 2 | + | + | + | + |
| | 3 | + | + | + | + |
| | 4 | + | + | + | + |
| | 5 | + | + | + | + |
| - | 6 | 0 | 0 | 0 | 0 |

Wilt bacteria were not found in plants that were not showing any symptoms. This PCR method can be used in epidemiological research, breeding lucerne for resistance to Cmi, and diagnostic phytosanitary laboratories to quickly identify Cmi.

Conclusion

According to the findings of our research, mycorrhizal symbioses (MPs) serve as selective microhabitats in terrestrial ecosystems for a diverse spectrum of fungal species. These MPs attract communities that are distinct from those found in the surrounding soil. This finding demonstrates that some forms of fungi are better suited to thrive on plastic surfaces than others, and that extensive colonization of the plastisphere may be achievable. As a consequence of this, non-natural fungal communities may develop in the soils of different regions of the world due to the presence of rubbish made of plastic. The MP serves as a host for fungus-causing illnesses and even accumulates them in terrestrial systems, including the environment immediately surrounding humans and Sub-Saharan Africa in particular. This is something that we have shown to be true. In plastisphere mycobiomes that were dominated

by diseases belonging to the Dothideomycetes and Tremellomycetes classes, respectively, a cryptococcal yeast and Phoma-like flamentous fungi were discovered. Findings similar to these suggest that these pervasive and extremely persistent pollutants may act as direct sources of infection and may also open up new pathways for infection by, for example, increasing pathogen loads and pathogen vectoring, which may potentially increase the risk of disease in both wildlife and humans. Researchers in the future should investigate the ecological and epidemiological repercussions of the most likely global disasters, and governments should address the possibility that plastic litter poses a threat to human health.

Reference

- [1].Sudarsono, Sudarsono, et al. "Pathogen Causing Phalaenopsis Soft Rot Disease 16S RDNA and Virulence Characterisation." Plant Protection Science, vol. 54, no. No. 1, 24 Nov. 2017, pp. 1–8, 10.17221/18/2017-pps. Accessed 11 Mar. 2020.
- [2].Pagter, Elena, et al. "Differences in Microplastic Abundances within Demersal Communities Highlight the Importance of an Ecosystem-Based Approach to Microplastic Monitoring." Marine Pollution Bulletin, vol. 160, Nov. 2020, p. 111644, 10.1016/j.marpolbul.2020.111644.
- [3]. Yusof Shuaib Ibrahim ; Ruthra Rathnam ; Sabiqah Tuan Anuar ; Wan Mohd, A. W. M. K. "ISOLATION and CHARACTERISATION of MICROPLASTIC ABUNDANCE in Lates Calcarifer from SETIU WETLANDS, MALAYSIA." Malaysian Journal of Analytical Science, vol. 21, no. 5, 23 Oct. 2017, 10.17576/mjas-2017-2105-07.
- [4].Maughan, H., et al. "Bacterial DNA Uptake Sequences Can Accumulate by Molecular Drive Alone." Genetics, vol. 186, no. 2, 13 July 2010, pp. 613–627, www.zoology.ubc.ca/~redfield/PDF/Maughan2010_1.pdf, 10.1534/genetics.110.119438.
- [5].Gopalakrishnan, V., and S. Muniraj. "Neem Flower Extract Assisted Green Synthesis of Copper Nanoparticles Optimisation, Characterisation and Anti-Bacterial Study." Materials Today: Proceedings, vol. 36, 2021, pp. 832–836, 10.1016/j.matpr.2020.07.013
- [6].Brear, Paul, et al. "Synthesis and Structural Characterisation of Selective Non-Carbohydrate-Based Inhibitors of Bacterial Sialidases." ChemBioChem, vol. 13, no. 16, 15 Oct. 2012, pp. 2374–2383, 10.1002/cbic.201200433.
- [7].Green, Benjamin C., and Charlotte L.E. Johnson. "Characterisation of Microplastic Contamination in Sediment of England's Inshore Waters." Marine Pollution Bulletin, Dec. 2019, p. 110788, 10.1016/j.marpolbul.2019.110788.
- [8].Bryxiová, M., and V. Kůdela. "Detection of Bacterial Wilt Pathogen of Lucerne by PCR." Plant Protection Science, vol. 38, no. SI 2 - 6th Conf EFPP 2002, 31 Dec. 2017, pp. 294–296, 10.17221/10472-pps.
- [9].Nanson, Jeffrey D., and Jade K. Forwood. "Structural Characterisation of FabG from Yersinia Pestis, a Key Component of Bacterial Fatty Acid Synthesis." PLOS ONE, vol. 10, no. 11, 5 Nov. 2015, p. e0141543, 10.1371/journal.pone.0141543.
- [10]. "Review of Phosphocholine Substituents on Bacterial Pathogen Glycans: Synthesis, Structures and Interactions with Host Proteins." Molecular Immunology,

vol. 56, no. 4, 31 Dec. 2013, pp. 563–573, www.sciencedirect.com/science/article/pii/S0161589013004057, 10.1016/j.molimm.2013.05.237.

- [11]. Tanner, Kaitlyn, et al. "The Bacterial Pathogen-Ubiquitin Interface: Lessons Learned FromShigella." Cellular Microbiology, vol. 17, no. 1, 28 Nov. 2014, pp. 35– 44, 10.1111/cmi.12390.
- [12]. Ricci, Ada, et al. "Visualizing the Relevance of Bacterial Blue- and Red-Light Receptors during Plant-Pathogen Interaction." Environmental Microbiology Reports, vol. 7, no. 5, 7 Aug. 2015, pp. 795–802, 10.1111/1758-2229.12320.
- [13]. Mathalon, Alysse, and Paul Hill. "Microplastic Fibers in the Intertidal Ecosystem Surrounding Halifax Harbor, Nova Scotia." Marine Pollution Bulletin, vol. 81, no. 1, Apr. 2014, pp. 69–79, 10.1016/j.marpolbul.2014.02.018.
- [14]. Galloway, Tamara S., et al. "Interactions of Microplastic Debris throughout the Marine Ecosystem." Nature Ecology & Evolution, vol. 1, no. 5, 20 Apr. 2017, www.nature.com/articles/s41559-017-0116, 10.1038/s41559-017-0116.
- [15]. Wingen, Lukas Martin, et al. "Versatile Synthesis of Pathogen Specific Bacterial Cell Wall Building Blocks." RSC Advances, vol. 12, no. 24, 2022, pp. 15046–15069, 10.1039/d2ra01915a.
- [16]. Yingkamhaeng, Naiyasit, et al. "Fabrication and Characterisation of Functionalised Superparamagnetic Bacterial Nanocellulose Using Ultrasonic-Assisted in Situ Synthesis." Fibers and Polymers, vol. 19, no. 3, Mar. 2018, pp. 489–497, 10.1007/s12221-018-7738-6.
- [17]. R.Manju, R.Manju. "Isolation, Identification, Characterisation of Bacterial Species from Keratinase Producing Microorganism." Indian Journal of Applied Research, vol. 1, no. 12, 1 Oct. 2011, pp. 178–180, 10.15373/2249555x/sep2012/63.