

EVALUATION OF MEDICINAL USES, PHYTOCHEMISTRY AND PHARMACOLOGICAL PROPERTIES OF *GREWIA OCCIDENTALIS* L. (MALVACEAE)

Alfred Maroyi

Department of Botany, University of Fort Hare, Private Bag X1314, Alice 5700, South Africa; Phone/Fax: 0027406022322; E-mail: amaroyi@ufh.ac.za

Abstract

Grewia occidentalis L. is a shrub or small tree widely used as traditional medicine in southern Africa. The current study critically reviewed the medicinal uses, phytochemistry and pharmacological properties of *G. occidentalis*. Results of the current study are based on literature survey conducted using various search engines such as Web of Science, Elsevier, Pubmed, Google scholar, Springer, Science Direct, Scopus, Taylor and Francis, and pre-electronic sources such as books, book chapters, scientific journals and other grey literature obtained from the University library. This study revealed that *G. occidentalis* used mainly to ease childbirth and hasten labour, as ethnoveterinary medicine, and traditional medicine for bladder ailments, impotence, infertility, respiratory infections, sores and wounds. Pharmacological research identified flavonoids, lipids, phenolics, tannins and triterpenoids from the bark, fruits, leaves, roots and wood of the species. The crude extracts of *G. occidentalis* and phytochemical compounds isolated from the species exhibited antibacterial, antigonococcal, antifungal, anti-HIV, anti-inflammatory, antioxidant and uterotonic activities. *Grewia occidentalis* should be subjected to detailed phytochemical, pharmacological and toxicological evaluations aimed at correlating its medicinal uses with its phytochemistry and pharmacological properties.

Keywords: Ethnopharmacology, *Grewia occidentalis*, herbal medicine, indigenous knowledge, Malvaceae, southern Africa, Tiliaceae

1. Introduction

Grewia occidentalis L. (Figure 1) is a shrub or small tree belonging to the Malvaceae family or the mallows. Evidence from recent molecular analysis has established that *G. occidentalis* belongs to the expanded Malvaceae family as opposed to its previous association with the Tiliaceae family [1-3]. Recent research has resulted in expansion of the family Malvaceae to include families such as Bombacaceae, Sterculiaceae and Tiliaceae [1-3]. The family Malvaceae consists of approximately 250 genera and 4200 species which are mostly herbs or shrubs [4,5]. The genus *Grewia* L. is a pantropical genus with about 300 recognised species of trees, shrubs and lianas distributed throughout the Old World [1]. The genus name *Grewia* is in honour of Nehemiah Grew (1641-1712) an English botanist, physician, author and microscopist, who is considered to be among the founders of plant anatomy as a branch of scientific research [6-8]. The species name *occidentalis* means “from the west” [6-8]. The synonyms of *G. occidentalis* include *G. chirindae* Baker.f., *G. microphylla* Weim. and *G. rudatisii* Burret [9,10]. The English common names of *G. occidentalis* include “assegai wood”, “cross berry”, “four corner”, “bow wood”, “button wood”, “dew berry”, “pink donkey berry” and “star flower”.



Figure 1: *Grewiaoccidentalis*: A branch showing leaves, flowers and fruits (photo: BT Wursten)

Grewiaoccidentalis is an evergreen to semi-deciduous multi-stemmed shrub or small tree with tangled crown that can grow to a height of 10 metres [7]. *Grewiaoccidentalis* sometimes occurs as a climber in closed evergreen forest and has a tendency to scramble. The bark of *G. occidentalis* is smooth and grey to grey-brown in colour. Leaves of *G. occidentalis* are simple and alternate, lanceolate to ovate in shape, light green and thinly textured but smooth with three prominent veins from the leaf base [7]. The flowers are pink to dark mauve in colour, showing yellow stamens in central mass, occurring in clusters and situated opposite the leaves. The fruit of *G. occidentalis* is reddish brown in colour, slightly fleshy and shiny drupe when mature, four-lobed and characteristically square in shape. *Grewiaoccidentalis* has been recorded in a variety of habitats such as mountain slopes, thornveld, forest margins, open woodland, bushveld, thicket, wooded grassland, arid highlands, arid Karoo, riverine bush, coastal dune bush and evergreen montane forests [11]. *Grewiaoccidentalis* has been recorded in Eswatini, Lesotho, Mozambique, South Africa and Zimbabwe at an altitude ranging from sea level to 2075 m above sea level [10]. The branches and bark which contain gum and tannin are tough, strong and fibrous and used to make string or rope [12,13]. *Grewiaoccidentalis* is variable and divided into two varieties, namely *G. occidentalis* var. *occidentalis* occurring in Zimbabwe and South Africa, and *G. occidentalis* var. *littoralis* Wild confined to coastal and sand dunes in Mozambique [9]. The leaves of *G. occidentalis* are browsed by game and livestock [14]. The ripe fruits are eaten by humans, often collected and dried for later use [7,13,15-17]. The leaves of *G. occidentalis* are used as leafy vegetables in South Africa [15,16,18-21].

Roots of *G. occidentalis* are an ingredient of a herbal mixture known as “isihlambezo” used to induce or augment labour and as postnatal medication to expel the afterbirth, and often administered to animals to expel the placenta and treatment of endometritis [12,22-24]. The “isihlambezo” ingredients include roots of *G. occidentalis*, *Agapanthus africanus* (L.) Hoffmans (roots), *Callilepislareola* DC. (roots), *Cliviaminiata* (Lindl.) Bosse (leaves), *Combretumerythrophyllum* (Burch.) Sond. (roots), *Crinum* spp. (bulb), *Gomphocarpusfruticosus* (L.) W.T. Aiton (roots), *Gunneraperpensa* L. (rhizomes), *Gymnanthemumcorymbosum* (Thunb.) H. Rob. (roots), *Pentanisiaprunelloides* (Klotzsch) Walp. (roots), *Rhoicissustridentata* (L.f.) Wild & R.B. Drumm. subsp. *cuneifolia* (Eckl. & Zeyh.) Urton (roots), *Scadoxuspuniceus* (L.) Friis & Nordal (bulb) and *Typhacapensis* (Rohrb.) N.E.Br. (rhizome) [12,22-24]. It is therefore, within this context that this study was undertaken aimed at reviewing the medicinal uses, phytochemistry and pharmacological properties of *G. occidentalis*.

2. Materials and methods

Several electronic databases were searched which included Web of Science, Elsevier, Pubmed, Google scholar, Springer, Science Direct, Scopus, Taylor and Francis. Additional information was obtained from pre-electronic sources such as books, book chapters, scientific journals and other grey literature obtained from the University library. The relevant terms *Grewiaoccidentalis* was paired with keywords such as “medicinal uses of *Grewiaoccidentalis*”, “phytochemicals of *Grewiaoccidentalis*”, “biological activities of *Grewiaoccidentalis*”, “pharmacological properties of *Grewiaoccidentalis*”, “ethnobotany of *Grewiaoccidentalis*”, and various other synonyms and common names of the plant species. The ultimate goal of this search was to explore articles that investigated the medicinal uses, phytochemical and biological activities of *G. occidentalis*.

3. Results and discussion

3.1 Medicinal uses of *Grewiaoccidentalis*

The bark, leaf, root and twig decoctions or infusions of *G. occidentalis* are mainly used to ease childbirth and hasten labour, as ethnoveterinary medicine, and traditional medicine for bladder ailments, impotence, infertility, respiratory infections, sores and wounds (Table 1; Figure 2). Research by De Wet and Ngubane [25] showed that the roots of *G. occidentalis* are mixed with those of *Rhoicissusdigitata* (L.f.) Gilg & M. Brandt, *Crotalaria monteiroi* Taub. ex Baker f., *Brideliacathartica* Bertol., *Garcinia livingstonei* T. Anderson and *Commiphoraneglecta* I. Verd. to cleanse blood, as postpartum, and traditional medicine for infertility and menstrual problems. Research by Robert and Robert [13] showed that the bark and twigs of *G. occidentalis* are mixed with leaves of *Centellaasiatica* (L.) Urb. as traditional medicine for ear problems, insect and tick bites, scratches and wounds. Similarly, research by Muthaphuli [26] and Semenya and Maroyi [2] showed that the bark of *G. occidentalis* mixed with bark of *Cassia abbreviata* Oliv. and *Ficussycomorus* L. as traditional medicine for diabetes mellitus. According to Dold and Cocks [28], the leaves of *G. occidentalis* are mixed with those of *Olea europaea* L. subsp. *africana* (Mill.) P.S. Green and *Zanthoxylumcapense* (Thunb.) Harv. and the sap of *Aloe ferox* Mill. as ethnoveterinary medicine for gallsickness in stock.

Table 1: Medicinal uses of *Grewia occidentalis*

Medicinal use	Parts used and preparation	Country	Reference
Bladder ailments	Rootbark	South Africa	[8,29-36]
Blood cleansing	Roots mixed with those of <i>Rhoicissus digitata</i> (L.f.) Gilg & M. Brandt, <i>Crotalaria monteiroi</i> Taub. ex Baker f., <i>Brideliacathartica</i> Bertol., <i>Garcinia livingstonei</i> T. Anderson and <i>Commiphora neglecta</i> I. Verd.	South Africa	[25]
Boils	Roots	Lesotho	[30,37,38]
Charm and rituals	Stem and wood	Lesotho and South Africa	[39-42]
Diabetes mellitus	Bark mixed with bark of <i>Cassia abbreviata</i> Oliv. and <i>Ficussycomorus</i> L.	South Africa	[26,27]
Ear problems	Bark and twigs which are mixed with leaves of <i>Centella asiatica</i> (L.) Urb.	South Africa	[13]
Ease childbirth and hasten labour	Leaves, roots and twigs	South Africa	[6,8,9,13,15,30,31,33,35,36,43-47]
Impotence	Leaves, roots and twigs	Eswatini and South Africa	[6,9,13,15,36,44,46,48,49]
Infertility	Leaves, roots and twigs	Eswatini and South Africa	[6,9,13,15,36,43-46,48,50,51]
Infertility	Roots mixed with those of <i>R. digitata</i> , <i>C. monteiroi</i> , <i>B. cathartica</i> , <i>G. livingstonei</i> and <i>C. neglecta</i>	South Africa	[25]
Insect and tick bites	Bark and twigs which are mixed with leaves of <i>C. asiatica</i>	South Africa	[13]
Menstrual problems	Roots mixed with those of <i>R. digitata</i> , <i>C. monteiroi</i> , <i>B. cathartica</i> , <i>G. livingstonei</i> and <i>C. neglecta</i>	South Africa	[25]
Postpartum	Leaves, roots and twigs	South Africa	[13]
Postpartum	Roots mixed with those of <i>R. digitata</i> , <i>C. monteiroi</i> , <i>B. cathartica</i> , <i>G. livingstonei</i> and <i>C. neglecta</i>	South Africa	[25]
Respiratory infections (sore throat and tuberculosis)	Roots	South Africa	[52-56]
Scratches	Bark and twigs which are mixed with leaves of <i>C. asiatica</i>	South Africa	[13]
Sexually transmitted infections (syphilis and venereal diseases)	Roots	South Africa	[15,34,57]
Sores and wounds	Bark, leaves and twigs	Eswatini, Lesotho and South Africa	[6-9,12,13,21,30,32,33,35-38,44,47-49,58-61]
Wounds	Bark and twigs which are mixed with leaves of <i>C. asiatica</i>	South Africa	[13]
Tonic	Roots	South	[49]

		Africa	
Ethnoveterinary medicine (anthelmintics, gallsickness, heartwater and wounds)	Bark, leaves and twigs	South Africa	[62-66]
Gallsickness in stock	Leaves mixed with those of <i>Olea europaea</i> L. subsp. <i>africana</i> (Mill.) P.S. Green and <i>Zanthoxylum capense</i> (Thunb.) Harv. and the sap of <i>Aloe ferox</i> Mill.	South Africa	[28]

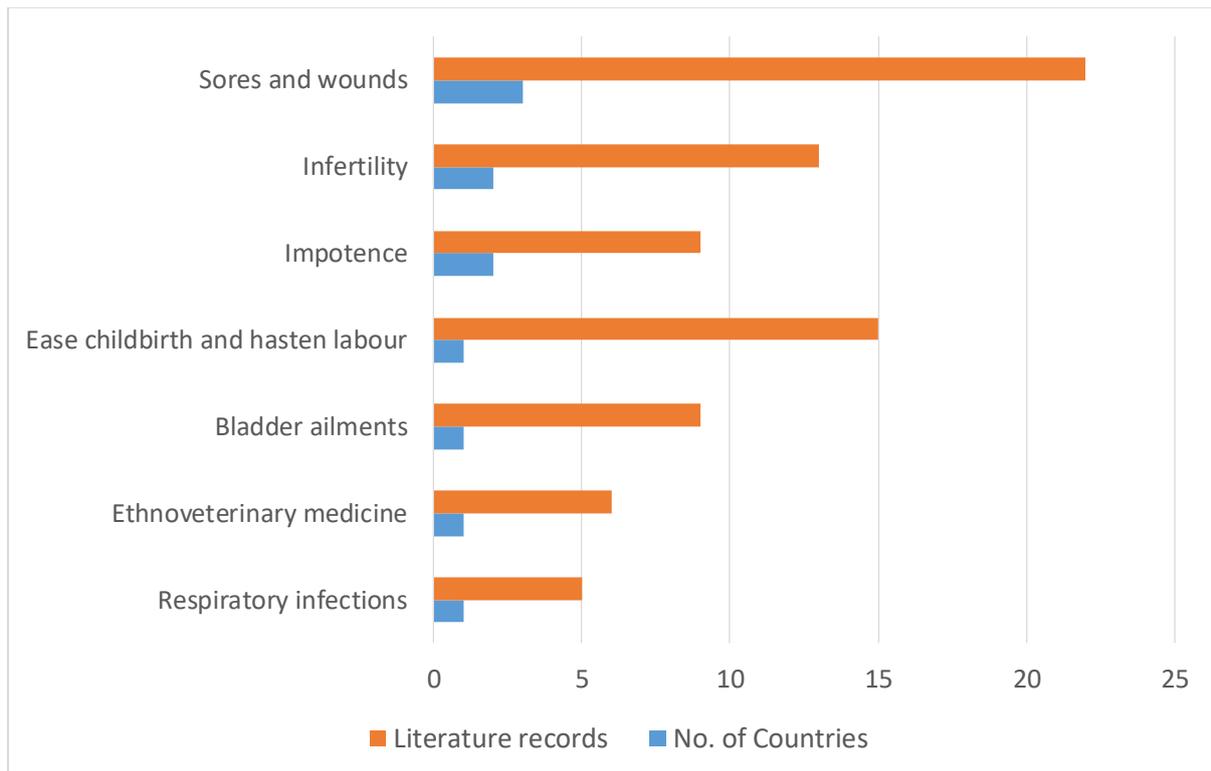


Figure 2: Medicinal uses of *Grewia occidentalis* in southern Africa

3.2 Nutritional and phytochemistry of *Grewia occidentalis*

Some researchers identified nutritional elements and phytochemical compounds from the bark, fruits, leaves, roots and wood of *G. occidentalis* (Table 2). The phytochemical compounds identified from the bark, fruits, leaves, roots and wood of *G. occidentalis* include flavonoids, lipids, phenolics, tannins and triterpenoids (Table 2). Some of these phytochemical compounds may be responsible for the pharmacological activities associated with *G. occidentalis*.

Table 2: Nutritional and phytochemistry of *Grewia occidentalis*

Nutritional and chemical compound	Value	Plant part	Reference
Acid detergent fibre (%)	27.0	Leaves	[67]
Ash (%)	12.7	Leaves	[67]
Calcium (mg/100g)	183.0	Leaves	[19]
Condensed tannins (% LCE) ^a	0.2	Roots	[57]
Coniferaldehyde ((E)-4-hydroxy-3-methoxy-cinnamaldehyde)	-	Bark and wood	[31]
Copper (%)	10.0	Leaves	[67]
Crude fibre (%)	14.4	Leaves	[67]

Crude protein (%)	6.0	Leaves	[67]
Flavonoids (µg CAE/g) ^b	1.2	Roots	[57]
Gallotannin (µg GAE/g) ^c	13.2	Roots	[57]
Iron (%)	187.5	Leaves	[67]
Lipid (%)	2.6	Fruits	[68]
Magnesium (mg/100g)	130.0	Leaves	[19]
Manganese (%)	96.0	Leaves	[67]
Neutral detergent fibre (%)	32.8	Leaves	[67]
Oleanonic acid	-	Bark and wood	[31]
Phosphorous (mg/100g)	76.0	Leaves	[19]
Potassium (mg/100g)	579.0	Leaves	[19]
Protein (%)	2.6	Fruits	[68]
Sinapaldehyde ((E)-4-hydroxy-3,5-dimethoxycinnamaldehyde)	-	Bark and wood	[31]
Sodium (mg/100g)	78.0	Leaves	[19]
Syringaldehyde (4-hydroxy-3,5-dimethoxybenzaldehyde)	-	Bark and wood	[31]
Total phenolics (mg GAE/g) ^c	11.7	Roots	[57]
Vitamin C (mg/100g)	18.4	Leaves	[19]
Zinc (mg/100g)	1.1	Leaves	[19]

^aValues expressed as percentage leucocyanidin equivalents (LCE) per gram plant extracts

^bValues expressed as catechin equivalents (CTE) per gram of plant extracts

^cValues expressed as gallic acid equivalent (GAE) per gram of plant extracts

3.3 Pharmacological properties of *Grewiaoccidentalis*

The following biological activities have been reported from the leaves, roots and shoots of *G. occidentalis* as well as phytochemical compounds isolated from the species: antibacterial [34,57,59,69,70], antigonococcal[34,57], antifungal [34,57], anti-HIV [34,57], anti-inflammatory [34,71], antioxidant [33,72] and uterotonic[31] activities.

3.3.1 Antibacterial activities

Grierson and Afolayan [59] evaluated the antibacterial activities of acetone, methanol and water extracts of *G. occidentalis* shoots against *Bacillus cereus*, *Bacillus pumilus*, *Bacillus subtilis*, *Micrococcus kristinae*, *Staphylococcus aureus*, *Enterobacter cloacae*, *Escherichia coli*, *Klebsiellapneumoniae*, *Pseudomonas aeruginosa* and *Serratiamarcescens* using the agar diffusion method. The methanol extract exhibited activities against all tested pathogens with minimum inhibitory concentration (MIC) values ranging from 1.0 mg/ml to 4.0 mg/ml [59]. Pretorius et al. [69] evaluated the antibacterial activities of crude extract of *G. occidentalis* leaves at a concentration of 50 mg/ml against *Clavibactermichiganense* sp. *michiganense*, *Agrobacterium tumefaciens*, *Erwiniacarotovora* sp. *carotovora*, *Xanthomonascampestris* sp. *phaseoli* and *Pseudomonas solanacearum* using the agar diffusion method with dimethyl dodecyl ammonium chloride as a positive control. The extract exhibited activities against *Pseudomonas solanacearum* with inhibition zone of 7.0 mm which compared favourably with inhibition zone of 12.0 mm exhibited by the positive control [69]. Mulaudzi [34] and Mulaudzi et al. [57] evaluated the antibacterial activities of ethanol, dichloromethane, water and petroleum ether extracts of *G. occidentalis* roots against *Escherichia coli*, *Staphylococcus aureus*, *Klebsiellapneumoniae* and *Bacillus subtilis* using the microdilution method with neomycin (0.1 mg/ml) as a positive control. The extracts exhibited activities against the tested pathogens with the MIC values ranging from 0.8 mg/ml to >12.5 mg/ml [34,57]. Lall et al. [70] evaluated antibacterial activities of ethanol extract of *G. occidentalis* leaves against *Mycobacterium smegmatis*, *Mycobacterium tuberculosis* and *Propionibacterium acnes* using micro-dilution technique with ciproflaxin and tetracycline as positive controls. The extract exhibited weak activities against tested pathogens with MIC values ranging from 500.0 µg/mL to >1000.0 µg/mL in comparison to MIC values of 0.3 µg/mL and 0.8 µg/mL exhibited by the positive controls ciproflaxin and tetracycline, respectively [70].

3.3.2 Antigonococcal activities

Mulaudzi [34] and Mulaudzi et al. [57] evaluated the antigonococcal activities of ethanol, dichloromethane and petroleum ether extracts of *G. occidentalis* roots against *Neisseria gonorrhoeae* through determination of clear

zones of inhibition with ciprofloxacin as a positive controls. The extracts exhibited moderate activities with percentage inhibition ranging from 44.0% to 53.0% [34,57].

3.3.3 Antifungal activities

Mulaudzi[34] and Mulaudzi et al. [57] evaluated the antifungal activities of ethanol, dichloromethane, water and petroleum ether extracts of *G. occidentalis* roots against *Candida albicans* using the microdilution method with amphotericin B as a positive control. The extracts exhibited activities against the tested pathogen with MIC and minimum fungicidal concentration (MFC) values ranging from 1.6 mg/ml to 6.3 mg/ml and 3.1 mg/ml to 6.3 mg/ml, respectively [34,57].

3.3.4 Anti-HIV activities

Mulaudzi[34] and Mulaudzi et al. [57] evaluated the anti-HIV activities of methanol and water extracts of *G. occidentalis* roots against a non-radioactive HIV-1 reverse transcriptase colorimetric ELISA kit with combivir and kaletra as positive controls. The extracts exhibited activities with inhibition percentage of 55.0% at 1.0 mg/ml and half maximal inhibitory concentration (IC₅₀) values ranging from 0.9 mg/ml to 0.1 mg/ml, which were comparable to IC₅₀ values of 0.06 mg/ml to 0.3 mg/ml exhibited by the positive control [34,57].

3.3.5 Anti-inflammatory activities

Mulaudzi[34] and Mulaudzi et al. [71] evaluated the anti-inflammatory activities of aqueous, dichloromethane, 80% ethanol and petroleum ether extracts of *G. occidentalis* roots against the cyclooxygenase (COX-1 and COX-2) enzymes. The dichloromethane and petroleum ether extracts exhibited activities towards both COX-1 and COX-2 with percentage inhibition of at least 70.0% [34,71].

3.3.6 Antioxidant activities

Steenkamp et al. [33] evaluated the antioxidant activities of aqueous and methanol extracts of *G. occidentalis* bark by assessing their ability to scavenge the hydroxyl radical (HO•) as measured by means of electron spin resonance spectrometry with the spin trap α -phenyl-N-tert-butyl nitron. For water and methanol extracts, the hydroxyl radical scavenging activities were 58.0% and 78.%, respectively [33]. Moustafa et al. [72] evaluated the antioxidant activities of methanol extracts of *G. occidentalis* leaf using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging assay with ascorbic acid as a positive control. The extract exhibited free radical scavenging activities of 94.7% and half maximal effective concentration (EC₅₀) value of 11.9 μ g/ml which was comparable to EC₅₀ value of 4.1 μ g/ml exhibited by the positive control [72].

3.3.7 Uterotonic activities

Mulholland et al. [31] evaluated the uterotonic activities of aqueous extract and an off-line supercritical extract of *G. occidentalis* using the uterine muscle tissue of a pregnant guinea pig. Research by Mulholland et al. [31] revealed that the uterotonic activities exhibited by the species were due to the compounds oleanonic acid and coniferaldehyde ((E)-4-hydroxy-3-methoxycinnamaldehyde).

4. Conclusion

The present review summarizes the medicinal uses, phytochemistry and pharmacological properties of *G. occidentalis*. Based on presented information, there is not yet enough data correlating the medicinal uses of the species with its phytochemical and pharmacological properties. Detailed studies on phytochemical, pharmacological properties, toxicological properties, *in vivo* and clinical research involving both extracts and compounds isolated from the species are required.

Conflict of interest

No conflict of interest is associated with this work.

References

1. Bayer C, Kubitzki K. Malvaceae. In: Kubitzki K, Bayer C (Eds.), Flowering plants, dicotyledons. Berlin: Springer-Verlag; 2003, pp. 225-311.
2. Heywood VH, Brummitt RK, Culham MA, Seberg RGO. Flowering plant families of the world. Ontario: Firefly books Ltd; 2007.
3. Angiosperm Phylogeny Group. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Bot J Linn Soc* 2009;161:105–121.
4. Simpson MG. Plant systematics. Singapore: Academic Press; 2010.
5. Christenhusz MJM, Byng JW. The number of known plants species in the world and its annual increase. *Phytotaxa* 2016;261:201–217.
6. Palmer E, Pitman P. Trees for southern Africa covering all known indigenous species in Republic of South Africa, South West Africa, Botswana, Lesotho and Swaziland. Cape Town: AA Balkema; 1972.
7. Venter F, Venter J-A. Making the most of indigenous trees. Pretoria: Briza Publications; 2015.
8. Schmidt E, Lotter M, McClelland W. Trees and shrubs of Mpumalanga and Kruger National Park. Johannesburg: Jacana Media; 2017.
9. Palgrave, M.C. Keith Coates Palgrave trees of southern Africa. Cape Town: Struik Publishers; 2002.
10. Germishuizen G, Meyer NL. Plants of southern Africa: An annotated checklist. Pretoria: Strelitzia 14, National Botanical Institute; 2003.
11. Manning J, Goldblatt P. Plants of the Greater Cape Floristic Region 1: The core Cape flora. Pretoria: Strelitzia 29, South African National Biodiversity Institute, 2012
12. Watt JM, Breyer-Brandwijk MG. The medicinal and poisonous plants of southern and eastern Africa. Edinburgh: E and S Livingstone Ltd; 1962.
13. Roberts, M., Roberts, S. Indigenous healing plants. Briza Publications, Pretoria; 2017.
14. Van Wyk B, Van Wyk P. Field guide to trees of southern Africa. Cape Town: Struik Nature; 2013.
15. Mabogo DEN. The ethnobotany of Vhavenda. MSc Dissertation. Pretoria: University of Pretoria; 1990.
16. Magwede K, Van Wyk B-E, Van Wyk AE. An inventory of Vhavenda useful plants. *S Afr J Bot* 2019;122:57–89.
17. Van Wijk Y, Rust R, Uithaler EM, Wurz S. Ethnobotanical research at Klasies River linking past, present, and future. *Ethnobot Res Appl* 2019;18:1-24.
18. Liengme CA. Plants used by the Tsonga people of Gazankulu. *Bothalia* 1981;13:501-518.
19. Steyn NP, Olivier J, Winter P, Nesamvuni C. A survey of wild, green, leafy vegetables and their potential in combating micronutrient deficiencies in rural populations. *S Afr J Sci* 2001;97:276–278.
20. Bvenura C, Afolayan AJ. The role of wild vegetables in household food security in South Africa: A review. *Food Res Int* 2015;76:1001–1011.
21. Mokganya MG, Tshisikhawe MP. Medicinal uses of selected wild edible vegetables consumed by Vhavenda of the Vhembe District Municipality, South Africa. *S Afr J Bot* 2019;122:184–188.
22. Veale DJH, Furman KI, Oliver DW. South African traditional herbal medicines used during pregnancy and childbirth. *J Ethnopharmacol* 1992;36:185–191.
23. Varga CA, Veale DJH. Isihlambezo: Utilization patterns and potential health effects of pregnancy-related traditional herbal medicine. *Social Sci Med* 1997;44:911–924.
24. Maroyi A. From traditional usage to pharmacological evidence: Systematic review of Gunnera perpersa L. *Evidence-Based Compl Alt Med* 2016;vol 2016, article ID 1720123.
25. De Wet H, Ngubane SC. Traditional herbal remedies used by women in a rural community in northern Maputaland (South Africa) for the treatment of gynaecology and obstetric complaints. *S Afr J Bot* 2014;94:129–139.
26. Muthaphuli N. Ethnomedicinal survey of plant species used in the management of sugar diabetes by the Vhavenda people of Muraga village in Vhembe District. BSc Honours Dissertation. Thohoyandou: University of Venda; 2016.
27. Semanya SS, Maroyi A. A review of plants used against diabetes mellitus by Bapedi and Vhavenda ethnic groups in the Limpopo province, South Africa. *Asian J Pharmaceut Clinical Res* 2019;12:44-50.

28. Dold AP, Cocks ML. Traditional veterinary medicine in the Alice district of the Eastern Cape Province, South Africa. *S Afr J Sci* 2001;97:375–379.
29. Gerstner J. A preliminary checklist Zulu names of plants with short notes. *Bantu Stud* 1939;13:131–149.
30. Hutchings A, Scott AH, Lewis G, Cunningham AB. *Zulu medicinal plants: An inventory*. Pietermaritzburg: University of Natal Press, 1996.
31. Mulholland DA, Sewram V, Raynor M, Thornell K and Raidoo DM. Coupling SFE to uterotonic bioassay: An on-line investigation of the uterotonic activity of compounds from *Grewia occidentalis* (Tiliaceae). *S Afr J Bot* 2002;68:68–71.
32. Grace OM, Prendergast HDV, Jäger AK, Van Staden J. Bark medicines used in traditional healthcare in KwaZulu-Natal, South Africa: An inventory. *S Afr J Bot* 2003;69:301–363.
33. Steenkamp V, Grimmer H, Semano M, Gulumian M. Antioxidant and genotoxic properties of South African herbal extracts. *Mutation Res* 2005;581:35–42.
34. Mulaudzi RB. Pharmacological evaluation of medicinal plants used by Venda people against venereal and related diseases. PhD Thesis. Pietermaritzburg: University of KwaZulu-Natal; 2012.
35. Lall N, Kishore N. Are plants used for skin care in South Africa fully explored? *J Ethnopharmacol* 2014;153:61–84.
36. Gulumian M, Yahaya ES, Steenkamp V. African herbal remedies with antioxidant activity: A potential resource base for wound treatment. *Evidence-Based Compl Alt Med* 2018, article ID 4089541.
37. JacotGuillarmod A. *Flora of Lesotho*. Cramer: Lehre, 1971.
38. Moffett R. *Sesotho plant and animal names and plants used by the Basotho*. Bloemfontein: Sun Press, 2010.
39. Moteetee A, Van Wyk B-E. The medical ethnobotany of Lesotho: A review. *Bothalia* 2011;41:209–228.
40. Tshikalange TE, Mophuting BC, Mahore J, Winterboer S, Lall N. An ethnobotanical study of medicinal plants used in villages under Jongilanga tribal council, Mpumalanga, South Africa. *Afr J Trad Compl Alt Med* 2016;13:83-89.
41. Moteetee A. A review of plants used for magic by Basotho people in comparison with other cultural groups in southern Africa. *Indian J Trad Knowl* 2017;16:229–234.
42. Moteetee A, Moffett RO, Seleteng-Kose L. A review of the ethnobotany of the Basotho of Lesotho and the Free State province of South Africa (South Sotho). *S Afr J Bot* 2019;122:21–56.
43. Bryant AT. *Zulu medicine and medicine-men*. Cape Town: C Struik, 1966.
44. Turner S. *Grewia occidentalis* L.; 2008. Available at: <http://www.plantzafrica.com/plantaefg/grewiaoccident.html>, accessed on 15 February 2021.
45. Abdillahi HS, Van Staden J. Application of medicinal plants in maternal healthcare and infertility: A South Africa perspective. *Planta Med* 2013;79:591–599.
46. Bhat RB. Medicinal plants and practices of Xhosa people in Transkei region of Eastern Cape, South Africa. *Indian J Trad Knowl* 2014;13:292–298.
47. Sagbo II, Mbeng WO. Plants used for cosmetics in the Eastern Cape province of South Africa: A case study of skin care. *Phcog Rev* 2018;12:139-156.
48. Long C. *Swaziland's Flora: siSwati Names and Uses*. Swaziland National Trust Commission; 2005. Available at: <http://www.sntc.org.sz/index.asp>, accessed on 15 February 2021.
49. Mhlongo LS, Van Wyk BE. Zulu medicinal ethnobotany: New records from the Amandawe area of KwaZulu-Natal, South Africa. *S Afr J Bot* 2019;122:266-290.
50. Steenkamp V. Traditional herbal remedies used by South African women for gynaecological complaints. *J Ethnopharmacol* 2003;86:97–108.
51. Amusan OOG. Ethical and environmental issues in bioprospecting for drugs through traditional medicine: The case of Swaziland. *Afr J Plant Sci Biotechnol* 2008;2:1-9.

52. Semenya SS, Maroyi A. Medicinal applications of plants by Bapedi traditional healers for sore throat and related symptoms in the Limpopo province, South Africa. *Med Plants Int J Phytomed Related Ind*2018;10:261-280.
53. Semenya SS, Maroyi A. Ethnobotanical survey of plants used by Bapedi traditional healers to treat tuberculosis and its opportunistic infections in the Limpopo province, South Africa. *S Afr J Bot* 2019;122:401–421.
54. Semenya SS, Maroyi A. Source of plants, used by Bapedi traditional healers for respiratory infections and related symptoms in the Limpopo province, South Africa. *J Biol Sci* 2019;19:101-121.
55. Semenya SS, Maroyi A. Source, harvesting, conservation status, threats and management of indigenous plant used for respiratory infections and related symptoms in the Limpopo province, South Africa. *Biodiversitas*2019;20:790-811.
56. Semenya SS, Maroyi A. Ethnobotanical survey of plants used totreat respiratory infections and related symptoms in the Limpopo province, South Africa. *J Herbal Med* 2020;24:100390.
57. Mulaudzi RB, Ndhkala AR, Kulkarni MG, Finnie JF, Van Staden J. Antimicrobial properties and phenolic contents of medicinal plants used by the Venda people for conditions related to venereal diseases. *J Ethnopharmacol* 2011;135:330–337.
58. Grierson DS, Afolayan AJ. An ethnobotanical study of plants used for the treatment of wounds in the Eastern Cape, South Africa. *J Ethnopharmacol* 1999;67:327–332.
59. Grierson DS, Afolayan AJ. Antibacterial activity of some indigenous plants used for the treatment of wounds in the Eastern Cape, South Africa. *J Ethnopharmacol* 1999;66:103–106.
60. Mabona U, Van Vuuren SF. Southern African medicinal plants used to treat skin diseases. *S Afr J Bot* 2013;87:175–193.
61. Sagbo IJ, Mbeng WO. Are plants used in the Eastern Cape province for cosmetics fully commercialized? *Indian J Pharmacol* 2019;51:140-149.
62. McGaw LJ, Eloff JN. Ethnoveterinary use of southern African plants and scientific evaluation of their medicinal properties. *J Ethnopharmacol* 2008;119:559–574.
63. Soyelu OT, Masika PJ. Traditional remedies used for the treatment of cattle wounds and myiasis in Amatolabasin, Eastern Cape province, South Africa. *Onderstepoort J Vet Res* 2009;76:393–397
64. Maphosa V, Masika PJ. Ethnoveterinary uses of medicinal plants: A survey of plants used in the ethnoveterinary control of gastro-intestinal parasites of goats in the Eastern Cape province, South Africa. *Pharm Biol* 2010;48:697-702.
65. McGaw LJ, Famuyide IM, Khunoana ET, Aremu AO. Ethnoveterinary botanical medicine in South Africa: A review of research from the last decade (2009 to 2019). *J Ethnopharmacol* 2020;257:112864.
66. Mthi S, Rust J, Yawa M, Tyasi L. Ethnoveterinary medicinal plants application for the treatment of tick-borne diseases in cattle around the Eastern Cape province of South Africa. *J Med Plants Econ Develop* 2020;4:a100.
67. Mthi S, Rust JM, Morgenthal TL. Partial nutritional evaluation of some browser plant species utilized by communal livestock in the Eastern Cape province, South Africa. *Appl Animal Husbandry Rural Develop* 2016;9:25-30.
68. Wilson A-L, Downs CT. Fruit nutritional composition and non-nutritive traits of indigenous South African tree species. *S Afr J Bot* 2012;78:30–36.
69. Pretorius PC, Magama S, Zietsman PC. Growth inhibition of plant pathogenic bacteria and fungi by extracts from selected South African plant species. *S Afr J Bot* 2003;69:186-192.
70. Lall N, de Canha MN, Reid A, Oosthuizen CB, Langhansova L, Mahore J, Winterboer S, Hamilton C, Kumar V, Gasa N, Twilley D. Antibacterial and anticancer activity of ethnomedicinal plants used in the Jongilanga community, Mpumalanga. *Int J Pharmacol. Phytochem Res* 2018;10:25–37.

71. Mulaudzi RB, Ndhala AR, Kulkarni MG, Finnie JF, Van Staden J. Anti-inflammatory and anti-mutagenic evaluation of medicinal plants used by the Venda people against venereal and related diseases. *J Ethnopharmacol* 2013;146:173–179.
72. Moustafa SMA, Menshawi BM, Wassel GM, Mahmoud K, Mounier MM. Screening of some wild and cultivated Egyptian plants for their free radical scavenging activity. *Int J PharmTech Res* 2014;6:1271-1278.