

NOVEL CARRIER FOR TRANSDERMAL PATCHES USED IN TRANSFEROSOMES: A REVIEW

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Abstract

Transferosomes deliver specialized transdermal medicines. These liposomes' edge contains phosphatidylcholine and an activator. Phospholipid vesicles are also used as transdermal drug carriers. The skin is the body's biggest organ, weighing three kilograms and covering 1.5 to 2.0 square meters. Liposomes, noisomes, and microemulsions are drug carriers used in transdermal drug administration that do not adequately transport drugs through the skin. Using the principle of rational membrane design, we recently created a new composite body called Transfersomes. Transfersomes can pass through the stratum corneum's smaller-than-normal pores into the viable skin beneath it. This is because it may be malformed. In vitro measurements of vascular vesicle size, entrapment efficiency, distortion, and density are possible. The "osmotic gradient" penetrates the stratum corneum intracellularly or transcellular. Transferosomes have various advantages, including biocompatibility and biodegradability. Some advantages include oxidative degradation and expensive cost. The transfersomes were made using rotary evaporation sonication. There are phospholipids, surfactants, and medicine. The transfersome's size distribution and zeta potential can be utilized to assess its performance. Vesicle morphology, vesicle density, entrapment efficiency Drug content turbidity measurement bending or permeability grading ability to overcome obstruction Surface charge and density In vitro drug release from skin Body balance. Transferosomes can be used for transdermal vaccination, targeted subcutaneous delivery, and controlled release.

Keywords: Transfersomes, Transdermal drug delivery system, Modified Transfersomes, Entrapment

Introduction

These are the most extensively utilized routes of drug delivery, with oral administration being favored for most small-molecule drugs. Oral administration is good for dose, mobility, and patient autonomy. For these reasons, oral medication is still the most convenient. Oral administration of most therapeutic peptides and proteins is unsuccessful due to stomach breakdown and epithelial size limitation. Injecting macromolecules has several disadvantages, including the intrusive nature of the process, lower patient acceptance/compliance rates, and the necessity for a competent administrator. Transdermal medication administration may address some of the fundamental limitations of traditional drug delivery systems (TDD).

Transdermal Drug Delivery

TDD distributes medications systemically through healthy, unbroken skin. The drug passes through the stratum corneum, deeper epidermis, and dermis without collecting in the dermal layer. When medications reach this level, they are absorbed via cutaneous microcirculation. TDD provides several advantages over other drug delivery systems. It is a non-invasive alternative to parenteral procedures. Transdermal absorption is achievable due to the skin's large surface area and ease of application. Hazardous side effects are less likely to develop because pharmacokinetic profiles are less variable and have lower peak activity. The lower dose frequency may benefit patients who are asleep, vomiting, or self-administering. Avoiding pre-systemic metabolism increases bioavailability. The presence of dendritic cells in both the epidermal and dermal layers of the skin makes TDD a prospective vaccine route for therapeutic proteins and peptides. A simple, needleless system like TDD for immunization has been the focus of several researchers due to the need for an affordable and non-invasive method in nations like the imperfect world.

Transfersomes

The aqueous compartment and edge activator in transfersomes make them excellent for biological transport. Their aqueous core coated by a lipid bilayer makes these ultra-deformable vesicles self-optimizing and self-regulating. Because transfersomes are flexible, they can be compressed to a smaller size than a vesicle while maintaining their vesicle integrity. Unlike typical liposomes, which include natural (egg phosphatidylcholine—EPC) or synthetic (dipalmitoyl phosphatidyl glycerol—DPPG) phospholipids, transfersomes contain a single-chain surfactant as an edge activator. EAs collaborate with a lipid to improve penetration to form a combination that makes the transfersoma pliable and flexible. This is owing to EAs' exceptional ability to destabilize membranes. Transfersomes, which can infiltrate pores smaller than their own diameter, provide these advantages. Despite being fractured, the transfersomes maintained their diameter after passing through the smaller pores. This transfersomal formulation functions better than regular liposomes because of the presence of EAs. Solubilizing hydrophobic medicines with EAs can improve drug entrapment efficiency in transferomal formulations. Using EAs to solubilize and fluidize skin lipids can improve skin permeability. The effects of EAs associated with skin penetration vary depending on EA type and concentration. Surfactants are also employed as penetration enhancers and edge activators. Amphiphilic compounds have lipophilic alkyl chains with hydrophilic head groups. Anionic surfactants have a lower critical micelle concentration than cationic surfactants, and their skin penetration is improved. Unlike cationic and anionic, Nonionic surfactants have an uncharged polar head group. Nonionic surfactants are considered less harmful and less irritating than anionic surfactants. Solubilizers, emulsifiers, and potent P-glycoprotein inhibitors are among the various functions of these substances. These formulations are used in peripheral medication targeting, transdermal vaccination, and other "transdermal delivery" methods. Several studies have shown that transfersomes can transport bioactive chemicals and hydrophilic and lipophilic molecules through the skin at over 50% efficiency. A specific cell architecture prevents significant molecular interactions between the skin surface and its depth. Even water may penetrate the epidermal barrier at 0.4 mg/cm²/h. Transcutaneous penetration rates revealed that the transferome could move drug molecules through the skin more than 50% faster than the untrapped drug. The transfersomes could carry the drug molecules through the skin 50% faster than untrapped drug molecules. Transfersomes also outperform liposomes in transferring lipophilic fluorescent markers through mouse skin by over 50%. The vesicular structure of transfersomes contains lipophilic and hydrophilic components. Vesicles smaller than 600 nm cannot enter deeper skin layers, whereas vesicles smaller than 300 nm can. The skin's viable epidermal and dermal layers contain the most vesicle content, less than 70 nm. Skin penetration was also boosted with 120-nm transfersomes compared to larger ones. These new transfersome delivery technologies ensure excellent phytoactive transport, bioavailability, and stability in herbal compositions. As a result, herbal ingredients can be encapsulated in transfersomes and used therapeutically for skin, hair, and eye care. The ability of transfersomes to penetrate deep epidermal layers and transport medications into the systemic circulation has been demonstrated in numerous investigations. The transfersome could potentially revolutionize medicine delivery.

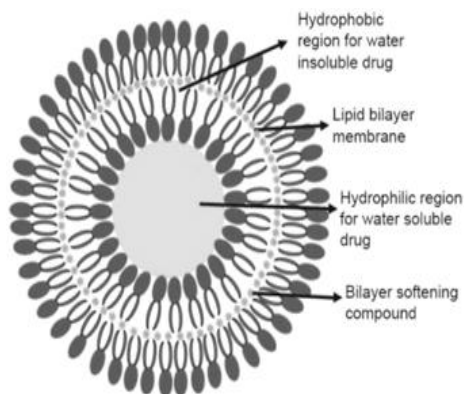


Fig 1: Transfersomes

Advantages of Transfersomes

- Transfersomes' hydrophobic and hydrophilic moiety structure allows them to bind a broad spectrum of medicinal substances. They can bend and pass through constrictions 5-10 times smaller than their own diameter without any loss.
- The system's deformability helps it to penetrate intact vesicles better. Some examples of low and high molecular weight drugs that can be carried are analgesics.
- Biocompatible and biodegradable due to the natural phospholipids used.
- Lipophilic medicines entrap over 90% of their targets.
- Some substances that protect medications from the breakdown in the body are proteins and peptides.
- They act as a depot and can be used for both systemic and topical pharmaceutical administration. They can be simply scaled up because the simple procedure excludes unnecessary or inappropriate ingredients.
- Liposomes appear to be the most similar structure to transfersomes. However, transfersomes outperform liposomes in terms of functionality because they are more malleable and flexible.
- Because the membrane is so flexible, transfersomes can squeeze through gaps as small as their own diameter.
- Transfersomes' hydrophobic and hydrophilic moiety structure allows them to bind to a broad spectrum of therapeutic substances.
- Transfersomes can flex and pass through a narrow constriction without injury (between 5 and 10 times smaller than their own diameter). Undamaged vesicles can easily pass through tight junctions due to their deformability. These carriers can transport analgesics, anesthetics, corticosteroids, sex hormone anticancer insulin, and gap junction proteins. Liposomes are made of natural phospholipids and are biodegradable. They are phospholipids. Lipophilic medicines are well-entrapped by them, at around 90% entrapment efficiency.

Disadvantages of Transfersomes

- Because of their susceptibility to oxidative destruction, transfersomes are highly unstable.
- Furthermore, natural phospholipids' purity is another factor in rejecting transfersomes as drug carriers.
- There is a price to pay for transfersome formulations.

Material for Transfersomes

It self-assembles into PC bilayers and then shuts, forming a self-adaptive and optimized mixed lipid aggregation to produce the transfersome and increased elasticity and permeability result from using a bilayer softener (such as a biocompatible surfactant or an amphiphile medication). When it comes to an "edge activator" in the system, this is the second component. Because it causes the lipid bilayer to become more fluid and elastic by disturbing its

structure, an edge activator is often utilized. A nonionic surfactant was used as the edge activator 30 in Van den Berg's elastic vesicles, which were introduced in 1998. Transferosomes can be made more flexible by using surface-active compounds. The resulting flexible and permeable transferosome vesicle may quickly adjust to the surrounding stress by modifying the concentration of each bilayer component at the bilayer. Transferosomes can follow the epidermis' natural water gradient if administered to patients in non-occlusive situations, reducing the likelihood of full vesicle rupture. Phospholipids (soya phosphatidylcholine, dipalmitoylphosphatidylcholine, etc.) make up most of these microcapsules, with surfactants (ethanol, methanol) making up the remaining 10 percent to 25 percent. A phosphate buffer solution (pH 6.5-7) is then used to moisten them. A dye, such as Rhodamine 123 or Nile red, is required for Confocal Scanning Laser Microscopy (CSLM).

Materials commonly used for the preparation of transferosomes are summarized in the table below:

Ingredient	Function	Examples
Phospholipid	Vesicle forming component	Soya Phosphatidylcholine Egg Phosphatidylcholine Disteryl Phosphatidylcholine
Surfactant	For providing flexibility	Sodium Cholate Sodium deoxy Cholate Tween 80 Span 80
Alcohol	As a solvent	Ethanol Methanol
Dye	For Confocal Scanning Lasser Microscopy(CSLM) Study	Rhodamine-123 Rhodamine-DHPE, Flurescein-DHPE
Buffering agent	As a hydrating medium	Saline phosphate buffer(pH 6.5) 7% v/v ethanol Tris buffer (pH 6.5)

Preparation of Transfersomes

1.

- A. Phospholipids and surfactants are dissolved in a volatile organic solvent (chloroform-methanol) to make a thin film. The organic solvent then evaporates at high temperatures (room temperature for pure PC vesicles, or 500C for dipalmitoylphosphatidylcholine). Overnight, a vacuum was employed to remove the remaining solvent.
- B. An hour of 60°C rotation hydrates a thin film of buffer (pH 6.5). Then they were removed after two hours of a swell time.
- C. This work used a bath sonicator or a 40°C probe to create tiny single-cell vesicles. This was done with a sandwich of 200 and 100 nm polycarbonate membranes.

B. The following steps were used to prepare transfersomes using a modified handshaking lipid film hydration approach.

1. All three components were dissolved in a 1:1 ethanol/chloroform combination. The organic solvent was evaporated by shaking the mixture at 43°C for some time. A thin film of lipids formed on the flask wall due to the spinning. Over the course of the following day, the solvent was allowed to evaporate.
2. The film was rehydrated for 15 minutes at the same temperature and phosphate buffer (pH 7.4). At temperatures ranging from 2 to 8 degrees Celsius, the transfersome suspension is fully hydrated in about an hour.

Evaluation of Transfersomes

- **Vesicle size distribution and zeta potential**

Malvern Zeta sizer was used to evaluate vesicle size, distribution, and zeta potential.

- **Vesicle morphology**

Both DLS and photon correlations spectroscopy can measure vesicle diameter. The size of the material was measured using photon correlations or raman scattering (DLS) on distilled liquid samples diluted with filtered saline and membrane filters. Transfersome vesicles can be observed via TEM, phase-contrast imaging, etc. A vesicle's stability can be assessed by tracking its growth and shape over time. DLS evaluates the average size and TEM structural changes.

- **No. of vesicles per cubic mm**

This critical metric is required for optimizing the composition and other process aspects. Parameter: Non-sonicated transfersome formulations are diluted five times with 0.9 percent sodium chloride solution. Further study can be done with an optical microscope and a hemocytometer. This is done using the formula: (Overall number of Transfersomes counted x dilution factor x 4000)/Total number of squares counted.

- **Entrapment efficiency**

The percentage of drug entrapment is how the entrapment efficiency is calculated. Mini-column centrifugation was used to separate the untrapped drug from the entrapped drug. Centrifugation was followed by either 0.1 percent Triton X-100 or 50 percent n-propanol disruption of the vesicles.

- **Drug content**

A modified phase high-performance liquid chromatography (HPLC) method can be used to evaluate the drug concentration of a pharmacopoeial medicament.

- **Turbidity measurement**

Nephelometers can be used to gauge the drug's turbidity in an aqueous solution.

- **Degree of deformability or permeability measurement**

Permeability testing is an essential and unique assessment of transfersomes. For their deformability testing, the scientists use clean water. To manufacture transfersomes, a large number of known-sized pores (through a sandwich of various microporous filters with pore diameters between 50 nm and 400 nm) are used (through a sandwich of different microporous filters with pore diameters between 50 nm and 400 nm, depending on the starting transfersomes suspension). After each pass, DLS measurements record particle size and size distributions.

- **Penetration ability**

The penetration ability of Transfersomes can be tested using fluorescence microscopy.

- **Occlusion effect**

In the case of classic topical medicines, occlusion of the skin may help penetration. However, elastic vesicles have the same issue. Hydrotaxis (movement towards the water) is the principal mechanism that drives vesicles from the skin's relatively dry surface to its water-rich deeper layers. Occlusion affects hydration forces by preventing skin evaporation.

- **Surface charge and charge density**

This tool determines the surface charge and density of Transfersomes.

- **In-vitro drug release**

The penetration rate is evaluated in vitro. Before doing costly in vivo tests, the formulation is optimized utilizing in vitro data such as time to steady-state permeation and steady-state permeation flux. To determine drug release, a tiny column centrifuge extracts free drug from a suspension of transfersomes cultivated at 320C. Drug released is determined by multiplying the initial drug entrapped by zero (100 percent entrapped and 0 percent released).

- **In-vitro Skin permeation Studies**

This investigation used a 2.50 cm² Franz diffusion cell adapted to hold 50 ml of the receiver compartment. Drug testing using goat skin in phosphate buffer (pH 7.4). (7.4) (7.4) The permeation test used butchered abdominal goat skin. The operation was completed by shaving and moisturizing the abdomen. We used cotton

swabs to remove the fatty tissue under the skin. It was kept between 0 and 40 degrees Celsius to preserve the skin.

Application of Transfersomes

❖ Delivery of proteins and peptides:

Transfersomes typically carry proteins and peptides. Oral peptides and proteins are large biogenic molecules that the body cannot absorb. This is why these peptides and proteins must still be injected into the body. Various solutions have been investigated. Subcutaneous administration of the same protein suspension has equivalent bioavailability to transfersomes. For example, adjuvant immunogenic bovine serum albumin in transfersomes is immunologically active after repeated epicutaneous therapy like the analogous injectable proteo-transfersomes preparations.

❖ Delivery of insulin

Transfersomes can transport large molecular weight drugs to the skin non-invasively. Insulin is commonly given subcutaneously, which is inconvenient. Insulin encapsulated in transfersomes (transfersulin) eliminates all issues. The first signs of systemic hypoglycemia appear 90-180 minutes after transfersulin injection on undamaged skin.

❖ Delivery of interferons:

Interferons, for example, are naturally occurring proteins with antiviral, antiproliferative, and immunomodulatory properties. Transfersomes as drug delivery devices may allow for controlled drug release and improved drug stability. Hafer et al. studied transdermal interleukin-2 and interferone formulations with transfersomes. They discovered that high quantities of transfersome-trapped IL-2 and INF- can be employed in immunotherapy.

❖ Delivery of corticosteroids:

Transfersomes can deliver corticosteroids. Transfersomes improve epicutaneous drug delivery site-specificity and overall drug safety. They are physiologically active at far lower concentrations than present skincare formulations.

❖ Transdermal immunization:

Another critical application of transfersomes is transdermal immunization, which employs transfersomes loaded with soluble protein like integral membrane protein, human serum albumin, and gap junction protein. These methods have at least two advantages: they can be used without injection, and they produce pretty high titers and, presumably, relatively high amounts of immunoglobulin A.

❖ Delivery of anesthetics:

Using anesthetics in transfersome suspension produces a topical anesthetic in under 10 minutes. The maximum ensuing pain insensitivity (80%) is similar to a subcutaneous bolus dose, although the effect of transfersomal anesthetics lasts longer.

❖ Delivery of NSAIDs:

NSAIDS have numerous GI adverse effects. Transdermal dispersion with ultra-deformable vesicles circumvents this. Diclofenac and Ketoprofen have been studied. Ketoprofen in a Transfersome formulation has been authorized for sale in Switzerland under the brand name Diractin. IDEA AG claims that further therapeutic medications based on Transfersome technology are currently in clinical studies.

❖ **Delivery of Anticancer Drugs:**

Transdermal administration of anticancer medications like methotrexate was investigated utilizing transfersome technology. The results were favorable. This opened up new possibilities for treating cancers of the skin, in particular.

❖ **Delivery of Herbal Drugs:**

Capsaicin Transfersomes have higher topical bioavailability than pure capsaicin since they can reach the skin barrier and provide nutrients locally.

Conclusion

An external shock can cause transfersomes to change shape fast and cheaply. These deformable particles can carry drugs past biological permeability barriers like skin with a standardized artificial system in a lab. Transfersomes can move through pores as small as 100 nm, almost as efficiently as water. Medication-loaded transfersomes can deliver up to 100mg of drug per hour through the skin (up to 100mg cm²h⁻¹). Although penetration of medication through the stratum corneum is a rate-limiting step, its principal limitations include the inability to carry larger size molecules.

For this reason, vesicular systems like Transfersomes were developed. The elastic vesicles deform to enter the skin via pores. Its formula is more efficient and safe than others. This delivery technology allows for controlled drug release. The disadvantages of conventional approaches can thus be avoided. Ultra-deformable vesicles may be the answer to transportation issues. They lack the rigidity of conventional vesicles and can transport even large molecules. They use a variety of processes to provide an excellent pharmaceutical carrier system. Transfersomes can pass through pores as small as 100 nm virtually as efficiently as water in simulated systems. Medication-loaded transfersomes can deliver up to 100mg of drug per hour through the skin (up to 100mg cm²h⁻¹). Drugs with little penetration due to undesired physicochemical qualities and those with a speedier and more specific action might all be delivered in this way.

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