

Applications of Nanoparticles in Cell and Molecular Biology

¹Dr. Kotte Shylaja

Assistant Professor in Chemistry (Humanities & Sciences)
St.Peters Engineering College,
Maisammaguda, Dhulapally, Medchal-Malkajigiri District, Secunderbad-500100,
India. Email : shylajasatyam@gmail.com

²Dr.R.R.Kumar

Assistant Professor, Department of Biochemistry,
Aarupadai Veedu Medical College and Hospital,
Vinayaka Missions Research Foundation, Puducherry
kumar.rangarajalu@avmc.edu.in

³Ms.S.Aruna, M.Sc.,M.Phil.,

Assistant Professor, Department of Biotechnology,
Marudhar Kesari Jain College for Women, Vaniyambadi.

Mail id - arunalakshita18@gmail.com

⁴Dr. Khushal N. Pathade

Assistant Professor and Head, P.G. Department of Botany
Dr. R. G. Bhoyar Arts, Commerce and Science College, Seloo Dist. Wardha, Maharashtra
Email- pathade.khushal@gmail.com

Abstract

Nanoparticles' customizable composition, size, shape, and surface chemistry make them ideal for biological applications. These created particles have opened new research vistas and helped answer basic biological issues. We shall discuss nanoparticle kinds and targeting concepts in this study. We will also discuss nanoparticles' use in medication delivery, imaging, sensing, and biological research.

Keywords: Nanotechnology; Sensing; Imaging; Nanoparticle; Drug delivery

Introduction

Nanotechnology is used in energy generation, industrial operations, and biological research. Nanotechnology has opened up many biological and medicinal research opportunities. Engineered nanoparticles (NPs) with different compositions and functions provide new tools and strategies in this field. NPs can detect analyses at attomolar concentrations and image biological processes at the cellular level. In this study, we examine NP kinds and their prospective uses in biology and biomedical research [1].

Different Nanoparticles

Size, shape, content, and functionality differ across NP platforms. Nanoprecipitation and lithography may be used to make each form of NP, notably polymeric ones. Although an in-depth investigation of NP platforms and fabrication techniques is beyond the scope of this publication, we will highlight the major traits and functions of each NP type that are important for biological research [2].

Oxidises Iron

Because of their super paramagnetic properties, super paramagnetic iron oxide nanoparticles (SPIONs) are used for passive and active imaging targeting. For stability, SPIONs have an iron oxide core covered with a hydrophilic material like dextran. Magnetite (Fe_3O_4) and maghemite (Fe_2O_3) cores are utilised in most SPIONs. These nanoparticles have size-dependent super paramagnetism, allowing magnetization in a magnetic field and loss of net magnetization when the field is removed. SPIONs work well as T2-weighted magnetic resonance (MR) contrast agents for cellular tracking and monitoring in biomedical research. SPIONs have lower toxicity, higher imaging sensitivity, and better imaging specificity than gadolinium-chelate contrast agents. SPION breakdown produces iron and iron oxide molecules that may be metabolised, retained in cells as ferritin, or integrated into haemoglobin. Two clinically authorised SPION agents for MRI are ferumoxides (120-180 nm) and ferucarbotran (60 nm). Molecular imaging applications like apoptosis detection and gene expression analysis use SPIONs. These nanoparticles may be functionalized with magnetic, optical, radioactive, and particular targeting ligands for multimodal imaging. SPIONs may also be used as medication delivery vehicles and non-invasive diagnostic techniques [3].

QDs, semiconductor particles with a diameter of less than 10 nm, were discovered in 1980. Nanoscale particles have size-dependent electrical and optical characteristics. The most researched QDs have a CdSe core and a ZnS cover. QDs emit light in a limited band yet absorb light broadly. QDs' brilliant colours are striking. They have extended lives, excellent quantum efficiencies, and photo bleaching resistance. Multiplexing is possible because QDs may be created with distinct biological specificities and stimulated and detected concurrently. Thus, QDs have significant benefits over organic fluorophore dyes for optical applications. QDs are used as fluorescent imaging tools in biological research for cell labelling and

biomolecule tracking. Their compact size makes them ideal for biological imaging and diagnostics. Due to their unique optical features and compatibility with biological systems, QDs have opened up new opportunities in various domains.

Gold

Gold nanoparticles (NPs) have unique optical and chemical characteristics, biocompatibility, and easy surface modification. Due to their unique interaction with light, gold NPs increase optical processes such light absorption, scattering, fluorescence, and SERS. Gold NPs may be used in biochemical sensing and detection, biological imaging, diagnostics, and therapeutics because to this characteristic. Gold NPs are used in colorimetric arrays to construct sensor platforms that use colour changes to detect analyses. Gold NPs serve as substrates in SERS, amplifying Raman scattering signals. Proteins and chemicals on the NP surface may be detected and identified by spectroscopy. Gold NP probes may also identify biomarkers linked to heart disease and cancer. Infrared phototherapy applications are promising because gold NPs transform absorbed light into heat. Localised heating of target tissues may aid therapeutic treatments. Gold nanoparticles are used in sensing, imaging, diagnostics, and therapeutic applications due to their unique optical and chemical characteristics, biocompatibility, and surface modification capabilities [4].

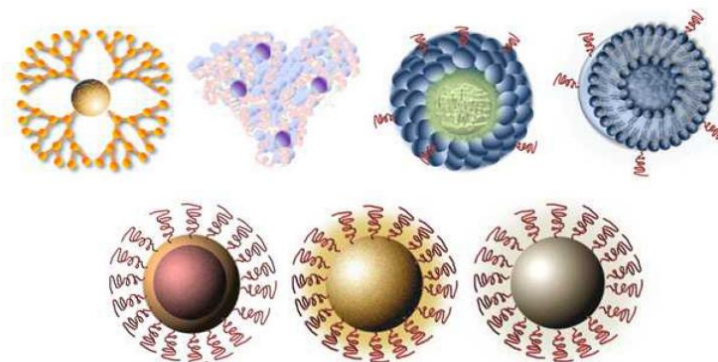


Fig.1 Diagram of nanoparticle kinds.

Targeted nanoparticles are a major topic in biological research. Targeted proteomics focuses on a subset of proteins to answer particular biological questions. In genetics, gene targeting in mice is the gold standard for mammalian gene function. Imatinib, trastuzumab, bevacizumab, and rituximab have revolutionised cancer therapy in drug development and molecular treatments.

Nanoparticles (NPs) have unique features that improve biological and biomedical research methods. Thus, targeting ideas in NP design are popular. Active or passive targeting may target NPs in vivo. Active targeting includes functionalizing NPs with ligands or antibodies that recognise and bind to target tissue-overexpressed cell surface receptors or molecules. Active targeting increases NP accumulation and absorption at targeted areas, improving medicinal effectiveness or diagnostic accuracy. Passive targeting uses leaky vasculature and poor lymphatic drainage to enhance NP accumulation in tumours. Small size, extended circulation, and adequate surface qualities enable NPs to preferentially concentrate in tumour tissues through the increased permeability and retention (EPR) effect. Targeting approaches in NP design have created interesting medicinal and biological research options. NPs may be designed to improve medication delivery, imaging, and therapy by using active and passive targeting techniques.

Passive Targeting

Passive targeting in NP systems has improved cancer treatment by using tumour tissue biology. Tumour vasculature is disorganised and irregularly branching. Solid tumours and inflammatory tissues have high vascular density, increased permeability, and poor lymphatic drainage. The increased permeability and retention (EPR) effect helps tumour tissue accumulate NPs. The EPR effect prolongs NP retention in tumour tissue, increasing concentrations relative to other tissues. NP composition, size, shape, and surface qualities determine passive targeting. NPs may target certain tissues or cells by carefully altering their physicochemical properties. Passive targeting using the EPR effect helps build NP systems that accumulate preferentially in tumour tissues. This approach uses the tumour microenvironment to improve treatment and reduce side effects. Researchers can improve passive targeting by altering NP characteristics, enhancing targeted medication delivery and cancer treatment [5].

Active Targeting

Targeting ligands improve NP system distribution to particular locations. These NP-conjugated ligands include small compounds, peptides, antibodies, antibody fragments, and nucleic acids like aptamers. Targeting ligands may be conjugated to NP surfaces covalently or noncovalently. Chemical conjugation attaches ligands to NP surfaces. Functional groups on the NP surface and the conjugated ligand interact to functionalize NPs. Targeting ligands

dictate conjugation methods. Peptides, antibodies, and their fragments are often conjugated to NPs via maleimide-thiol coupling. Maleimides react with sulfhydryl groups at slightly acidic pH to conjugate targeted ligands with thiol or cysteine residues. Carbodiimide-mediated amide coupling forms amide bonds between carboxyl groups and primary amines, another common technique. This method works for aptamers and small compounds with carboxylate or primary amine functional groups. Click chemistry processes may also conjugate functional groups to NPs with good yield and selectivity under moderate reaction conditions. This setting favours alkynes-azides [3+2] cycloaddition. These conjugation approaches may functionalize NP systems with targeted ligands to increase their selectivity and affinity for cells, tissues, and biomarkers. Active targeting directs NP systems to targeted areas, increasing medication delivery, diagnostics, and other biological uses [6].

Biomedical Nanoparticles

Nanoparticle sensors are widely used in sensor technologies. They excel at detecting low-concentration analytes, isolating pathogens, collecting cells, and monitoring molecular and cellular activities. These areas have created NP-based sensors. Analytes are detected. Receptors or recognition elements may selectively bind target analyses to NPs. This method lets NPs find disease-related biomarkers or genomic sequences. Quantum dots' adjustable fluorescence may boost analytical test signals. NP-based sensors detect and separate pathogens. Functionalized NPs, commonly coated with antibodies or aptamers, recognise and bind harmful bacteria and viruses. This allows sensors to identify pathogens in clinical samples quickly and sensitively. Magnetic NPs can isolate pathogens from complicated samples. Cell detection and capture depend on NPs. Functionalizing NPs with ligands that target cell surface indicators allows for cell detection and isolation. This method may identify uncommon circulating tumour cells for cancer diagnosis or isolate particular cell types for study or treatment. NPs monitor molecular and cellular activities. Fluorescent NPs that react to pH, ion concentrations, or enzymatic activity may visualise and measure molecular processes in live cells. Magnetic NPs are contrast agents in MRI to monitor cellular processes or therapeutic drug administration. Sensor development benefits from NPs' tiny size, broad surface area, and functionalization. Their excellent sensitivity, specificity, and multiplexing allow many biological target and process detection and monitoring applications [7].

Analyse Nanoparticles

Even at low concentrations, DNA, RNA, and protein detecting methods have improved. These revolutionary methods use nanoparticles' unique features. NPs are appropriate for surface ligand use to increase detection sensitivity or speed due to their huge surface area to mass ratio, tiny size, and composition-dependent properties. Targeting ligands help NPs bind and signal analyses, improving detection. NP biosensors use inorganic NPs, especially metallic or magnetic ones.

Pathogen-Detecting Nanoparticles

NP platforms with optical and magnetic characteristics have been studied for pathogen detection and isolation. Magnetic biosensors utilise antibodies-coated magnetic NPs to detect microorganisms. Researchers used this immunomagnetic technique in a new microfluidic device to draw molecules coupled to magnetic NPs from one flow stream to another using a local magnetic field gradient. This system separates biotinylated anti-*E. coli* antibodies and streptavidin-coated superparamagnetic iron oxide nanoparticles (SPIONs)-labeled *E. coli* from red blood cell solutions at blood densities. This approach may effectively separate pathogens in complicated samples[8].

Cell-Detecting / Separating Nanoparticles

Biological studies and applications need cell separation from complicated mixtures. Nanoparticles (NPs) can identify and trap low-frequency cell types. CTC detection and capture are significant uses. CTCs may reveal the biology of cancer metastasis and predict overall survival in metastatic breast, colorectal, and prostate cancer patients[9].

Nanoparticle Imaging

Nanoparticles (NPs) are new molecular imaging labels and contrast agents. NPs can monitor molecular targets and cellular responses connected to cancer and cardiovascular disorders using their unique features. This might improve molecular imaging and disease processes[10].

Imaging Nanoparticles

Targeted imaging using nanoparticles (NPs) has several advantages. First, their huge surface area delivers many imaging agents, increasing imaging sensitivity. Second, NPs may

passively target tissues in vivo via the increased permeability and retention (EPR) effect or actively target areas where the molecular target is expressed to increase contrast agent concentration. Targeted NP build up improves imaging. NPs may also be used as in vivo imaging amplifiers due to their modifiability. Finally, NPs may carry several imaging agents for multimodality imaging, increasing their targeted imaging flexibility [11].

Pharmaceutical Nanoparticles

Getting siRNA to biological researchers RNA interference (RNAi) is essential in cell culture and live organisms for targeting mRNA destruction using particular double-stranded RNAs. RNAi is commonly used to study gene functions, although siRNA delivery may be difficult. Due to their huge size and negative charge, intracellular nucleases degrade them, exact targeting to the right cellular compartment is challenging, and they rapidly clear and are unstable in vivo. Thus, effective and biocompatible delivery mechanisms are needed to properly use RNAi [12].

Subcellular Injections

Research is focused on efficiently delivering agents to organelles. Effective delivery requires accurate targeting and accessibility of medicinal and imaging agents to subcellular organelles. Delivering chemicals to certain organelles may reveal molecular processes that are yet unknown. Thus, improving drug distribution to subcellular organelles might improve therapeutic and imaging applications and our understanding of intracellular processes [13].

NPs' Biology

Understanding biological systems is essential to developing effective nanoparticles (NPs) for sensing, imaging, and medication administration. Thus, NP characteristics' biological implications are being studied. Size, shape, functional groups, surface charge, and composition affect NP interactions with biological systems in laboratories and live organisms. However, biological variables affecting NP behaviour require additional study. RES/MPS cells rapidly remove NPs. Researchers use "steal thing" to extend NP circulation in vivo by conjugating uncharged hydrophilic polymers like PEG to the NP surface [14]. However, PEGylated particles may induce PEG antibodies. To address this difficulty, one must understand NP-MPS bimolecular interactions. This understanding is essential for developing techniques to lengthen NP circulation durations in vivo while

minimising immunological reactions and antibody production. Thus, understanding NP interactions with biological systems, especially the complex MPS-NP connection, is crucial. Understanding will improve NP design, performance, and circulation times in vivo, resulting in better sensing, imaging, and drug administration applications[15].

Nanoparticle-Based Biological Research

Nanoparticles (NPs) have enabled new molecular and cellular biology studies. NPs influence cell signalling pathways, promote protein synthesis, and enhance methods. Quantum dots (QDs) are popular for their biological fluorescence applications. QDs have been studied for their use in immunostaining fixed cells and tissues, labelling membrane proteins, and visualising cytoskeleton filaments. For molecular imaging, these fluorescent probes are bright and photo stable. QDs have been used to study live cell molecules' dynamic behaviour [16].

EGFRs coupled to QDs may be visualised. Researchers tagged a tiny percentage of EGFR molecules with a compound of one CdSe QD and one anti-EGFR antibody Fab fragment (antiEGFR-Fab). Total internal reflection fluorescence microscopy allowed live cell molecular dynamics studies of EGFR-Fab-QD complexes. These advances show QDs and other NPs may help us comprehend biological processes. NPs' unique optical characteristics allow molecular studies of cellular activities, revealing complicated biological phenomena [17].

Conclusions and Proposed Research

Nanotechnology and nanoparticles (NPs) have many biological uses. They can image, sense, administer drugs, and analyse biological processes at the cell and biomolecule level. The growing interest in biological NP applications raises hopes for new and fascinating applications. Many essential genes and proteins have been found, but their assembly and integration are yet unknown. NP investigations may illuminate complicated signalling cascades and biological activities. NPs help researchers learn how biological components work together to complete tasks. NPs may help explain how cells react to mechanical forces and chemical signals in processes including apoptosis, cell division, and stem cell destiny [18].

NP-based monitoring and detection methods may reveal disease and injury metabolic pathways. NP sensors can detect biological substances that conventional tests cannot.

Targeted NP systems may bind or react to these molecules to enhance detection and molecular imaging. NPs can also solve biological research delivery difficulties such hydrophobic chemical solubility and stability in aqueous circumstances and nonspecific targeting. Nanoscale studies of biological systems, proteomics, genomics, and nanotechnology help us comprehend cellular function and processes. This information may inspire nanoscale techniques to restore cellular function in disorders. As nanotechnology approaches advance, biologists may more easily use NP methods and platforms in their study.

NPs used in fluorescence imaging, magnetic resonance imaging, and low-concentration analyse detection are commercially accessible. However, size, shape, functional groups, surface charge, and composition may affect NP interactions with biological systems. For biological applications, NP formulation toxicity must be carefully assessed. Chemists and nanotechnologists must standardise and simplify NP synthesis to fully use NP technology in biological research. Nanotechnology researchers and biologists must collaborate to find new applications and synergies. This integrative strategy will enhance NPs in biological research and provide breakthrough discoveries [19, 20].

References

1. Allhoff, F.; Lin, P.; Moore, D. What is nanotechnology and why does it matter? : from science to ethics. Chichester, UK ; Malden, MA: Wiley-Blackwell; 2010. p. xp. 293
2. Bangham AD. Liposomes: the Babraham connection. *Chemistry and Physics of Lipids*. 1993; 64:275–285. DOI: [http://dx.doi.org/10.1016/0009-3084\(93\)90071-A](http://dx.doi.org/10.1016/0009-3084(93)90071-A).
3. Torchilin VP. Recent advances with liposomes as pharmaceutical carriers. *Nat Rev Drug Discov*. 2005; 4:145–160.
4. . Felgner PL, Ringold GM. Cationic liposome-mediated transfection. *Nature*. 1989; 337:387–388.
5. Felgner PL, Gadek TR, Holm M, Roman R, Chan HW, Wenz M, Northrop JP, Ringold GM, Danielsen M. Lipofection: a highly efficient, lipid-mediated DNA-transfection procedure. *Proceedings of the National Academy of Sciences*. 1987; 84:7413–7417.
6. . Bangham AD. Liposomes-The Babraham Connection. *Chem. Phys. Lipids*. 1993; 64:275–285.

7. Hawkins MJ, Soon-Shiong P, Desai N. Protein nanoparticles as drug carriers in clinical medicine. *Advanced Drug Delivery Reviews*. 2008; 60:876–885.
8. Gradishar WJ, Tjulandin S, Davidson N, Shaw H, Desai N, Bhar P, Hawkins M, O'Shaughnessy J. Phase III Trial of Nanoparticle Albumin-Bound Paclitaxel Compared With Polyethylated Castor Oil–Based Paclitaxel in Women With Breast Cancer. *J Clin Oncol*. 2005; 23:7794–7803.
9. Harries M, Ellis P, Harper P. Nanoparticle Albumin–Bound Paclitaxel for Metastatic Breast Cancer. *Journal of Clinical Oncology*. 2005; 23:7768–7771.
10. . Gradishar WJ. Albumin-bound paclitaxel: a next-generation taxane. *Expert Opinion on Pharmacotherapy*. 2006; 7:1041–1053. [PubMed: 16722814]
11. Gref R, Minamitake Y, Peracchia M, Trubetskoy V, Torchilin V, Langer R. Biodegradable longcirculating polymeric nanospheres. *Science*. 1994; 263:1600–1603
12. Chan, J.; Valencia, P.; Zhang, L.; Langer, R.; Farokhzad, O. *Cancer Nanotechnology*. Grobmyer, SR.; Moudgil, BM., editors. Vol. 624. Humana Press; 2010. p. 163-175.
13. Torchilin VP. Micellar Nanocarriers: Pharmaceutical Perspectives. *Pharm Res*. 2007; 24:1–16.
14. Wang AZ, Gu F, Zhang L, Chan JM, Radovic-Moreno A, Shaikh MR, Farokhzad OC. Biofunctionalized targeted nanoparticles for therapeutic applications. *Expert Opinion on Biological Therapy*. 2008; 8:1063–1070.
15. Medina SH, El-Sayed MEH. Dendrimers as Carriers for Delivery of Chemotherapeutic Agents. *Chemical Reviews*. 2009; 109:3141–3157.
16. Fréchet JMJ. Dendrimers and supramolecular chemistry. *Proceedings of the National Academy of Sciences*. 2002; 99:4782–4787.
17. Svenson S, Tomalia DA. Dendrimers in biomedical applications—reflections on the field. *Advanced Drug Delivery Reviews*. 2005; 57:2106–2129.
18. Lee CC, MacKay JA, Frechet JMJ, Szoka FC. Designing dendrimers for biological applications. *Nat Biotech*. 2005; 23:1517–1526.
19. Mintzer MA, Grinstaff MW. Biomedical applications of dendrimers: a tutorial. *Chemical Society Reviews*. 2011; 40:173–190.
20. Weissleder R. Molecular Imaging in Cancer. *Science*. 2006; 312:1168–1171.